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Impact of Stress on Obesity-Targetted Signaling Pathways for Novel Drug Discovery

Sujoy Kundu¹, Sumithra Mohan^{2*}, Chitra V³

^{1,2*,3}Department of Pharmacology, SRM College of Pharmacy, SRM Institute of Science and Technology, Kattankulathur-603203, Chengalpattu, Tamil Nadu, India

Corresponding Author: Dr. Sumithra Mohan^{2*}

^{2*}Ph.DAssociate ProfessorSRM College of Pharmacy, SRMISTKattankulathur-603203, Chengalpattu, Tamil Nadu, India Email ID: ^{2*}sumithrm@srmist.edu.in

Abstract

Obesity is an epidemic that affects millions of people across the world. It is a metabolic disorder that can be characterized by an excess buildup of adipose tissue. Stress can be defined as the organism's false response to any kind of demand or challenge to equilibrium. Whenever an organism comes in contact with any sort of stressor (physical, social, etc.), all the originating internal stress mechanisms are triggered to start a flight or fight response and thereby deal with the stress event. Several animal stress models are thought to exhibit unique metabolic traits, with some animals exhibiting signs of anorexia and a reduction in body weight, while others show an increase in food consumption and body weight and become more prone to disorders of metabolism. Hormones that are released by the endocrine organ gut, such as ghrelin, leptin, glucagon-like peptide-1 (GLP-1), peptide YY (PYY), pancreatic polypeptide (PP) as well as cholecystokinin (CCK), have quite an influence on the maintenance of homeostasis and energy balance by generating satiety and food termination. Leptin, a hormone that is released by adipocytes into the bloodstream is necessary for the regulation of body weight and the proper balance of energy. Obesity is caused primarily by leptin deficits or genetic abnormalities in the signaling of the leptin mechanism. A therapeutic approach by the use of probiotics as an external living system for the control of obesity is usually suggested by recent research that examined the relationship of the gut microbiota with obesity. The superfamily of mitogen-activated protein kinases (MAPK) includes the c-Jun N-terminal kinase (JNK) family. In obesity, JNK is a signal transducer that has been extensively studied. This review explores various kinds of rodent stress models and signaling pathways that affect metabolic outcomes to better understand stress-induced obesity.

Keywords: Obesity, Stress, Leptin, JNK, Nrf2 Pathways, Stress Model.

1. Introduction

The World Health Organization (WHO) reports that obesity incidence has increased twice since 1980. More people die from overweight or obesity than from underweight in most countries of the world ^[1]. The regulation of body weight and energy balance depends mainly on the GI tract, which is the body's largest endocrine organ and secretes a variety of gut hormones, which include cholecystokinin (CCK), ghrelin, peptide YY (PYY), as well as glucagon-like peptide (GLP-1) ^[2]. Obesity has become an epidemic, and it is now widely accepted. Some medical experts believe that obesity is the largest global cause of mortality that may be prevented ^[3]. In fact, according to some research, weight gain considerably reduces life expectancy, and those who are obese can pass away a decade or more earlier than people who are of normal weight ^[4]. Stress might be one of the major environmental factors that impact metabolism and food intake that leads to obesity.

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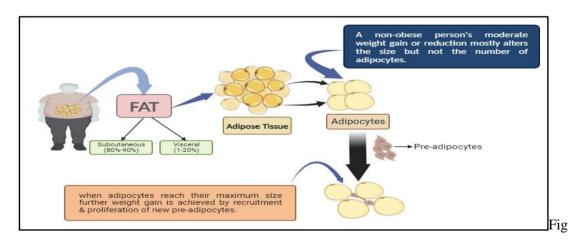
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The two main glucocorticoids that are secreted as a response to stressful conditions are cortisol and corticosterone in humans as well as rats, respectively. Glucocorticoids oppose insulin's efforts to store energy in numerous tissues by partially reducing insulin secretion ^[5, 6]. A transitory state of insulin resistance (IR) generated by glucocorticoids inhibits the production of hepatic glucose and regulates the transport of glucose to peripheral tissue, leading to a state of acute hyperglycemia^[7, 8].

This review focuses on the understanding of the pathophysiology and metabolic alterations linked with obesity and also the employment of animal models to study stress-related activities. Additionally, it also focuses on the signaling pathways that govern food consumption as well as the role of serotonin and homeostasis.

Pathophysiology of Obesity

The major fat organ, which is regarded as an endocrine organ is the adipose tissue (AT) that plays a part in the immune system's response ^[9]. The majority of the body's fat mass is made up of subcutaneous adipose tissue (80%) and visceral fat (20%). Triglycerides (TG) compose 90% of the lipids that make up the lean body mass, which contributes over 80% of the weight [Fig. 1] ^[10,11]. Based on homeostasis, AT may either carry out the lipolysis of TG by the enzyme lipoprotein lipase (LPL), which generates free fatty acids (FFA) as well as glycerol, or by generating acyl-CoA from extracellular fatty acids or glycerol-3-phosphate, it may esterify these compounds. Lipogenesis and glyceroneogenesis, respectively, are terms given to these processes ^[12–14]. As a result, over long periods of positive energy balance (when energy intake exceeds energy loss), AT works by engaging preadipocytes, which grow rapidly and undergo hypertrophy into full-grown adipocytes, resulting in vascular and stromal development.



1.Subcutaneous (80%–90%) and visceral (1%–20%) adipose tissue compose the majority of the body's fat mass, respectively. Adipose tissue is made up of adipocytes, and after adipocytes reach their maximal size, the recruitment and proliferation of new pre-adipocytes allow for further weight gain. AT induces the release of active biomarkers from stromal cells i.e. endothelial cells, fibroblasts, leukocytes, and macrophages, as well as, to a small amount, adipocytes. The proteins, hormones, and cytokines that are secreted are known to be adipocytokines or adipokines including leptin, adiponectin, resistin, visfatin, retinol-binding protein 4 (RBP-4), and many more [13–16]. Peripheral afferent system, central processing system, and efferent system control obesity pathogenesis. The peripheral processing system, divided into peripheral appetite-suppressing and peripheral appetite-stimulating systems, is suppressed by adipocytes secreting leptin and adiponectin [Fig 2]. The central processing system, divided into central appetite suppressing and central stimulating systems, is stimulated by the gut hormones ghrelin and obestatin. NPY, a central stimulus, decreases energy

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expenditure and increases food intake, maintaining energy balance. Recently, terms like "adiposopathy" and "diabesity" which denote the near association between T2DM and obesity, have been developed to describe the close link between the pathology of AT and metabolic disorders [17, 18]. The main sites in which IR occurs are the muscle, liver, AT, and kidney [19]. In the community, overweight and obesity are associated with problems that increase the death risk by 2–3 times in obese people [20].

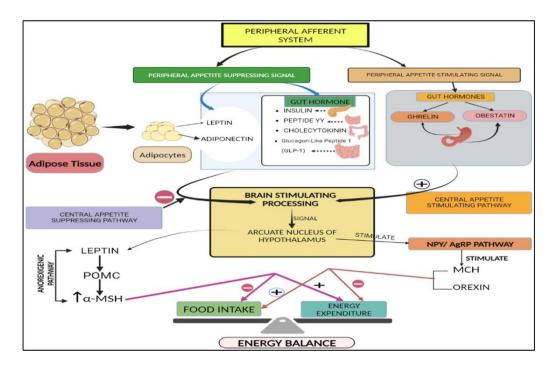


Fig 2. The peripheral afferent system, central processing system & efferent system control the pathogenesis of obesity. The peripheral processing system is divided into peripheral appetite suppressing & peripheral appetite stimulating systems. The peripheral suppressing system, through adipocytes secreting leptin & adiponectin, suppresses the central processing system. The peripheral stimulating signals including gut hormones ghrelin and obestatin, stimulate the central processing system. The central suppressing system, through α -MSH suppresses food intake and increases expenditure while the central stimulating system through NPY decreases energy expenditure and increase food intake maintaining energy balance. POMC: Proopiomelanocortin; α -MSH: α -Melanocyte-stimulating hormone; MCH: Melanin-concentrating hormone

Stress, Gut & Metabolic Hormones and Obesity

There are over 20 different hormones released by the gastrointestinal tract, which is the largest endocrine organ in the body. The main function of gut hormones is nutrition uptake, although they also perform other functions such as digestion and absorption ^[21]. These are a few of the most significant hormones-

• Peptide YY (PYY)

PYY along with pancreatic polypeptide (PP) are the neuropeptide Y (NPY) family members and the diverse system of the cell along the gut-brain axis express them. PYY is produced by a special type of endocrine cells of the distal gut and is termed PYY because it contains a tyrosine residue. In both the fed and fasted states, PYY3-36 is the predominant circulating hormone ^[22, 23]. With the intake of food, levels of PYY are elevated, reach their peak by 1-2 hours, and remain raised for an extended period, showing that it functions primarily as a component of satiety rather than being a terminator of food

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PYY mediates its effects through the Y2 receptor, which is found all over the CNS. The hypothalamic regulation of consumption of food by the brainstem and vagal afferents is affected by PYY3–36, and it may potentially directly affect this control. Recent research suggests that several brain regions except the homeostatic region may also affect hunger, which is mediated by PYY 36 [25]. Numerous cases have shown that PYY3-36 may reduce weight in part as a result of its effects on fuel partitioning and energy loss ^[26].

• Pancreatic polypeptide (PP)

The f cells of the pancreatic islets secrete PP, producing minimally in the distal portion of the gut. After food intake, the PP circulation increases for about 30 minutes and continues for minutes after the termination of the meal ^[27]. In contrast, somatostatin prevents PP secretion. PYY has a certain level that persists for an extended period after the meal has terminated, indicating that it may promote satiety. PP has a greater affinity for the Y4 receptor, even though it is thought that the brainstem regulates the majority of this hormone's anorectic actions ^[28]. Numerous earlier investigations found that repeated peripheral PP administration in animals decreased the intake of food. According to several human investigations, when normal-weight subjects are studied, their food intake is reduced by 25% ^[29-31]. No study has associated changes in body weight with fasting or postprandial PP ^[31, 32].

• Glucagon-LikePeptide 1 (GLP-1)

GLP-1, a member of the incretin hormone family, is a gut-derived hormone that stimulates the release of insulin in response to glucose, among other functions. Two forms of GLP-1 are produced by the intestinal L cells, GLP-11-37 and GLP-11-36 amide [33]. The major hormone in circulation is GLP-17-36. This hormone's secretion is mainly stimulated by food. GLP-1 acts on the GLP-1 receptor (GLP-1R), found both in peripheral tissues as well as the CNS. The hypothalamus and brainstem both express c-fos, which mediates GLP-1 activity [34]. The receptors of GLP-1 are usually found in the periphery and have a significant role in the termination of meals and the feeling of satisfaction [35]. GLP-1 administration in both the peripheral and central regions significantly reduced food intake by rodents [36]. Additionally, the satiated rat's food intake was doubled by antagonistic central GLP-1 receptors [37]. Although the majority of research has shown that GLP-1's concentration remains unchanged in both the overweight and lean subjects, there is enough proof to suggest that GLP-1 has a role in the pathophysiology of obesity.

• Cholecystokinin(CCK):

This is released with the help of specialized cells known as I cell. The tripeptidal peptidase II enzyme inactivates this hormone, which is formed by the posttranslational moderation of the procholecystokinin gene with a combining with peptides [38]. CCK levels rise biphasically for about 25 minutes after food intake and then remain elevated for about 3 hours. CCK-1 receptor mediate CCK's anorectic effects, according to genetic and pharmacological research. The primary CCK receptors are CCK-1 and CCK-2, which are both present in the brainstem and hypothalamus. Satiety is generated by CCK, when CCK-1 binds to the vagus nerve [39]. According to some findings, compared to obese or lean individuals, morbidly obese individuals had lower fasting levels. The effectiveness of CCK has recently demonstrated a limited level of anti-obesity activity. CCK appears to have a greater impact on satiety than satiation [40].

• Ghrelin

It is a 28-amino acid neuropeptide hormone produced by cleaving a larger precursor, preproghrelin. Ghrelin is a hormone generated from the gut that is produced by the X/A-like cells of the gastric oxyntic mucosa surrounding the stomach [41]. Ghrelin specifically promotes consumption of food by triggering a several of hypothalamic, brainstem, and midbrain nuclei [42-46]. The only known peptide between the gut and the brain that stimulates appetite is ghrelin. The binding of certain

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molecules of medium-chain fatty acids, particularly octanoate, is involved in this alteration. This acylation is carried out by the enzyme gastric o-acyl transferase (GOAT) [47]. The orphan GH-secretagogue receptor (GHS-R1a) is the endogenous ligand through which ghrelin exerts its effects on adiposity. The acyl ghrelin (active form) can only cross the blood-brain barrier (BBB), which also activates and binds to the GHS-R1a receptor [48]. According to several studies, the ARC of the hypothalamus is where ghrelin's control of appetite functions is centrally situated [49]. The regulation of ghrelin-induced food intake is mostly dependent on the vagus nerve and brainstem [50]. Like PYY3–36, ghrelin enables us to recognize truly satisfying meals. It has been shown that ghrelin administration (both centrally and peripherally) indirectly triggers hypothalamic corticotropin-releasing factor (CRF) neurons, leading to stimulation of the HPA axis [51]. In Prader-Willi syndrome, where ghrelin levels rise and obesity develops, the pathogenic involvement of ghrelin has been suggested. However, ghrelin has been shown to be inversely correlated with BMI in non-syndromic plasma [52,53].

• Leptin

The peripherally derived hormone leptin affects the consumption of food and homeostasis and is expressed according to the size and number of adipocytes. The metabolic effects of stress are also influenced by leptin.Once it is produced, leptin stimulates the ARC's anorecticPOMC/ Cocaine-amphetamine related transcript (CART) neurons and regulates the ability in which ghrelin and other metabolically active hormones may cause these neurons to become depolarized [54–56].According to one study, intraperitoneal administration of glucocorticoids (emulating acute stressor levels) reduced consumption of food and body weight by increasing concentrations of leptin [57]. When glucocorticoids were administered centrally, the same group showed that leptin had a different function since NPY expression increased along with food consumption and body weight [58].

• Glucocorticoids

Visceral obesity has been linked to increased HPA axis activity in both humans and rodents, as well as elevations in glucocorticoid concentrations that can have an impact on the brain and peripheral tissues ^[59]. As with other steroid hormones, lipophilic glucocorticoids cross the BBB and affect the production of hypothalamic peptides that regulateintake of food and homeostasis ^[60–62]. Numerous brain areas significantly associated with energy balance, including the ARC, LH, and PVN, have been shown to express glucocorticoid receptors (GRs) ^[63]. Due to their capacity to modify behaviors that affect energy intake and expenditure, glucocorticoids are thought to have a significant impact on the connection between obesity and stress^[64–66]. The relationship between glucocorticoid levels and obesity, however, might not be as clear-cut as previously assumed.In addition to stimulating, inhibiting, and interacting with other metabolically active hormones (such as insulin, leptin, CRF, and others), glucocorticoids play a significant role in controlling feeding behavior^[61, 67–69]. Every stressor can initiate the release of glucocorticoids. The HPA axis reaction and the response to food intake, however, differ depending on the stressor. The duration and intensity of the current stressor have a significant role in how much the HPA axis gets involved and, consequently, how much glucocorticoids are generated in response to the stressor ^[70–72].

Signaling Pathway of Stress-Mediated Obesity

Obesity is a disorder of metabolism characterized by an abundant buildup of adipose tissue and has been associated with several types of molecular signaling pathways. Chronic stress can lead to a specific kind of obesity known as stress-induced obesity. The following signaling pathways [Fig. 3] have been linked to stress-related obesity:

Leptin Signaling Pathway:

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Leptin receptor b (LEPRb) belongs to the interleukin 6 (IL-6)-type cytokine receptor family that has three domains: extracellular, intracellular, and a single membrane-spanning domain ^[73–75].LEPRb interacts with Janus Kinase 2 (JAK 2), although it lacks intrinsic enzymatic activity ^[73, 75]. On certain tyrosines, leptin triggers the activation of JAK2 and its autophosphorylation ^[76, 77]. Tyr 985, Tyr 1077, and Tyr 1138 are the three tyrosine residues of LEPRb that are phosphorylated by JAK2 ^[77–79].

LEPRb Tyr1138-emanated JAK2/STAT3 signaling

Leptin causes JAK2 to phosphorylate LEPRb on Tyr 1138. After that, phosphorylated Tyr 1138 then interacts with the SH2 domain of STAT3 (signal transducer and activator of transcript 3) [80]. JAK2 associated with LEPRb subsequently phosphorylates STAT3, leading to nuclear translocation and dimerization [81]. Suppressors of cytokine signaling 3 (SOCS3) and neuropeptides are two examples of target genes that STAT3 dimers regulate by acting as transcription factors in the nucleus [77, 82]. Genetic studies have shown that leptin can reduce obesity depending on the JAK2/STAT3 pathway [83,84].

LEPRb Tyr1077-emanated JAK2/STAT5 signaling

Activation of STAT5 is facilitated by JAK2, which phosphorylates Tyr1077 on LEPRb by leptin, which binds to the SH2 domain of STAT5 ^[85, 86]. The activation of STAT5 is also partially modulated by phospho-Tyr1138 ^[86]. In mice, STAT5 activation in hypothalamic neurons lower food consumption, but STAT5 elimination in the CNS result in the obesity and hyperphagia ^[87].

LEPRb Tyr985-emanated SHP2/ERK signaling

A binding site for the SH2 domain of protein tyrosine phosphatase 2 (SHP2) is produced by phospho-Tyr985. The extracellular signal-regulated kinase (ERK) pathway activated by leptin is mediated by SHP2 [88, 89]. Under some circumstances, SHP2 could also downregulate JAK2/STAT3 signaling [90]. Leptin's anti-obesity action acts through the SHP2 pathway, as demonstrated by the fact that mice with the SHP2 gene eliminated in the brain develop obesity at an early stage [91–93]. According to the study, the substitution of Phe for Tyrosine-985 results in the elimination of Tyr985 phosphorylation, which elevated obesity and diet-induced leptin resistance [94]. Unexpectedly, a further investigation showed that removing Tyrosine 985 phosphorylation prevents female mice from developing diet-induced obesity [95]. The outcomes of Tyr985's phosphorylation are probably influenced by the levels of intracellular SOCS3.

• PI3K/AKT SignalingPathway:

The phosphoinositide 3-kinase (PI3K) pathway is also required for leptin activation ^[84, 96]. Mice become obese when IRS2 is deleted from the brain ^[97, 98]. Leptin-induced anorexia in mice is prevented by pharmacologically inhibiting PI3K in the hypothalamus ^[96]. The stimulation of the PI3K pathway by leptin is mediated by the adaptor protein SH2B1, which has an SH2 domain and binds to both the IRS and JAK2 proteins ^[99]. Leptin resistance and obesity are caused by disruptions in the SH2B1 gene ^[100, 101].

The forkhead box O1 (FoxO1) signaling branch

PI3K/Akt is a significant downstream effector of FoxO1, a transcription factor important for nutritional energy balance ^[102]. Several sites on FoxO1 are phosphorylated by Akt, which causes FoxO1 to be retained in the cytoplasm and become inactive ^[103]. In mice, food consumption and body weight are reduced when FoxO1 is deleted from POMC neurons ^[104, 105].IRS2 null mice with removed FoxO1 are able to reverse obesity phenotypes, while LEPRb neurons with removed IRS2 become obese and have an energy imbalance ^[106].

The mammalian target of rapamycin complex 1 (mTORC1/ ribosomal S6 kinase (S6K) signaling branch

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The mTOR/S6K pathway is a further step in the PI3K/IRS pathway's downstream processing ^[76]. Leptin stimulates mTORC1 to activate, which subsequently phosphorylates and triggers S6K in the hypothalamus ^[107, 108].

• JNK Signaling Pathway

The JNK signaling pathway may be activated by obesity, alcohol, and radiation to modulate inflammation, proteins, and oxidative stress [109]. JNK can be activated by the MAP2K (MKK4 and MKK7) enzymes that are present upstream of JNK. Both MKK4 and MKK7 are capable of phosphorylating JNK concurrently at the threonine-183 and tyrosine-185 sites; however, MKK4 usually phosphorylates tyrosine whereas MKK7 phosphorylates threonine. Furthermore, several upstream MAP3K is controlled by a variety of different upstream factors, such as TNFα and FFA which in turn activate MKK4 and MKK7 [110-112]. JNK is known as c-Jun N-terminal kinase as it was first identified as a kinase that mainly phosphorylates only the nuclear transcription factor c-Jun. JNK moves into the nucleus from the cytoplasm when the JNK signaling pathway is triggered, where it transphosphorylates Ser63 and Ser73 in the c-Jun transcription factor's amino-terminal active domain to activate it. The c-Jun transcription factor binds to the transcription factor activator protein-1 (AP-1) at the gene promoter after activation, triggering the expression of proinflammatory genes and protein synthesis (e.g., TNF, IL-1, IL-6, and IL-8), which leads to impaired glucose tolerance as a result of obesity and insulin resistance [113-114]. A primary function of JNK activity is to regulate metabolism by phosphorylating its substrate in the JNK signaling pathway, and it plays an important role in disease occurrence. These findings collectively demonstrate that JNK strongly influences metabolism, especially in conditions associated with inflammation and obesity [115].

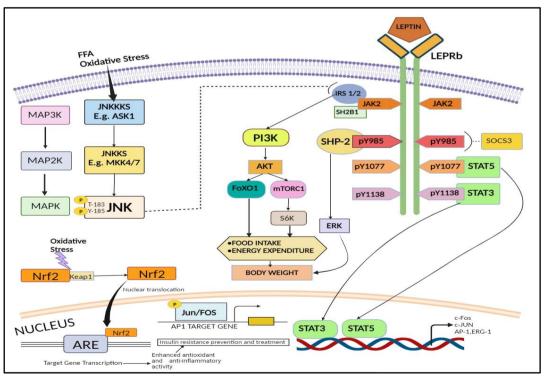


Fig. 3. The network of signaling pathways involved in stress mediated obesity.Nrf2: Nuclear factor erythroid 2-related factor 2, keap 1: kelch like ECH associated protein 1, IRS: Insulin Receptor Substrate, SH2B1: SH2B adaptor protein 1, pY985: tyrosine 985, pY1077: tyrosine1077, pY1138: tyrosine1138, ERK: extracellular signal-regulated kinase, S6K: ribosomal S6 Kinase

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• Nrf2 Signaling Pathway

When under minimal stress, Nrf2 is cytoplasmically bound to Keap1; however, when under oxidative stress, it dissociates from Keap1 and translocates to the nucleus. Following this, Nrf2 attaches to the antioxidant response element (ARE), which triggers the transcription of the genes expressing cytoprotective antioxidants. Due to this, increased antioxidant and anti-inflammatory activity has the potential to prevent or treat insulin resistance [116].

These pathways are not exhaustive, and stress-induced obesity may include other signalling pathways. However, these pathways provide insight into the complex mechanisms that underlie the onset of obesity and might potentially serve as therapeutic targets for obesity treatment.

Stress Models

• Visible Burrow System (VBS)

VBS was first designed as a realistic social stress agent to study agonistic actions in rodents ^[117, 118]. This model imitates a rat's natural surroundings by stimulating an underground burrow system. In the VBS, two adult female rats and four adult male rats are typically kept together. Within days, a social hierarchy forms, producing three subservient and one dominant maleanimal^[119]. Within the VBS, subservient animals have elevated initial corticosterone (CORT), elevated testosterone and consistently reduced ~10-15% of their body mass^[117, 120-124]. With other measures such as initial locomotor activity removed as contributing parameters, the reduction in body weight has been linked to changes in metabolic rate and decreased food intake ^[119, 124]. In the VBS, it has been shown that subservient animals produce lower levels of leptin and insulin, but the expression of orexigenic peptides and hormones is unchanged. Due to reduced food consumption and alteration in the rate of metabolism, subservient animals in the VBS show lower adiposity and a reduction in lean body mass ^[124]. The majority of endocrine and physiologic parameters are restored in animals who have been taken out of VBS and allowed to recuperate to enhance their metabolic phenotype, but they aren't restored to levels that are comparable to dominant or unstressed control animals in terms of body weight ^[119].

• Resident Intruder

The resident intruder model, also called as social defeat stress, has been used by researchers for a very long time to study how social dominance and subordination affect animals [125-127]. An animal is referred to as a resident when its territorial behavior toward its home cage quickly develops after being single-housed for ~1 week. The researcher introduces an entirely new, non-littermate rodent to the inhabitant within the home cage in the resident intruder model. Therefore, the new animal appears to be an invader within the cage and must compete within the social hierarchy for his position (usually held by males). While allowing for the development of a social hierarchy, it was demonstrated that keeping animals with their littermates shows no impact on anxious behavior or CORT generation in subservient animals and therefore is considered non-stressful [128]. In contrast, subservient rodent exhibit signs of stress while living with a new, non-mate rodent, including reduced immunological responses and increased food consumption, fat mass and body weight [128, 129-132].

Typically, there are two male rodents in this model- one intruder and one resident. Usually, the invader is placed in the resident's home cage, where animals are permitted to interact until one dominates another. A central barrier that divides the resident's home cage is used to separate the animals after interaction ^[128, 132]. This barrier allows the animals to exchange sensory information but restricts the use of physical contact. The experiment gives the animals a chance to establish a social hierarchy by removing the barrier and enabling them to interact, leading to the formation of four distinct groups: resident-dominants, resident-subordinates, intruder-dominants, and intruder-subordinate ^[128]. Some labs select new inmates every day of defeat in the context of persistent social defeat stress ^[130, 131, 133-131]

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^{138]}. The experimental individuals are presented with a novel social order throughout each distinct incident of defeat due to the deployment of novel inhabitants. For instance, in the resident intruder model, submissive animals have elevated CORT and blood glucose levels as well as elevated levels of orexigenic/adipogenic plasma ghrelin and elevated levels of hypothalamic NPY and Agouti-related protein (AgRP) expression ^[132, 137]. Ghrelin signaling increases lead to further increases in hypothalamic NPY/AgRP, which stimulate adiposity and consumption of food, with a focus on visceral adiposity, which causes obesity. It is capable of decreasing stress-mediated increases in food intake, body weight gain, as well as obesity when observed in submissive rodents by interrupting the signaling of ghrelin by pharmacological blockage or genetic manipulation ^[132].

• Chronic Mild Stressors (CMS)

CMS has been demonstrated to be capable of causing depression, along with the plethora of behavioral, physical, and neurochemical alterations that often present alongside depression (such as anhedonia) [139–141]. Chronic mild stress is caused by daily stresses. The animal may experience daily stressors such as living in a congested cage, a small room or room with wet bedding, tilting the room, shock in the foot, tail pinches, living in a filthy cage, predator smell, loud noise, changes to the cycle of light/dark (for example, 24 hours of light), and more [139–145]. The induction of anhedonia is the most notable physiological effect of CMS, but this paradigm is also able to impair established location preference, decrease libido, impair immunological action, and change the sleep cycle [140]. However, only a small number of CMS-based studies show that their experimental rodents are obese. However, CMS reduces the intake of enticing meals, which has an impact on body weight [142, 144, 145]. Before attempting to mimic human obesity, it is important to think about the types of CMS that may be used in human scenarios.

Physical Stressors

■ Foot shock/ Tail pinch

Rodent feeding behavior is influenced by two common physical stressors, such as foot shock and tail pinch. In this specific experiment, the availability of a highly palatable, calorie-dense food encouraged the rats to consume calories. Whether a tail pinch is enough to cause physiological alterations that may influence feeding behavior is unknown. The effect of general arousal is thought to be responsible for the feeding response rather than motivating feeding responses [146]. However, it has been demonstrated that foot shock increases the levels of leptin, insulin, and plasma glucose in the circulation of rats, indicating an anorectic hormonal profile [147, 149]. Animals exhibit no changes in food intake despite the anorectic hormonal profile imposed by repetitive foot shock, but they do maintain a positive energy balance and gain greater weight over time than controls not under stress [149]. According to the resident intruder paradigm, the metabolic effects of submissive animals occurred much more quickly than the body mass and body composition alterations caused by repetitive foot shocks [149].

Immobilization/ Restraint stress

Two more commonly used techniques for acute physical stress are immobilization and physical restraint. According to physiologic and neuroendocrine responses, restraint stress is referred to as mild stress, yet in the hypothalamus it has been shown to enhance the synthesis of orexigenic peptides [149]. Mild restraint stress can increase AgRP-expressing neurons in the ARC while simultaneously decreasing α-MSH receptors. Particularly acute restraint activates cFos in the lateral hypothalamus, paraventricular nucleus, and ARC; however, recurrent exposure diminishes the activation, indicating adaptation to the stress model [149-150]. On the other hand, immobilization stress is regarded as an effective physical stressor causing HPA axis hyperactivity since it significantly increases CORT secretion [151]. Following immobilization stress, it has been seen that rats eat less and acquire less

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weight, indicating evidence of the stressors' potency. Additionally, after immobilization stress, no alterations in NPY production are observed, providing hormonal support for the food intake responses to this kind of stressor model ^[152]. Data on the possibility of immobilization stress habituation are contradictory ^[152-154].

Cold stress

In the rodent and human stress paradigms, exposure to cold physical stressors has been employed. In humans, cold physical stressorscause cortisol levels to significantly increase 15 minutes after the initiation of the stressor [155]. The release of cortisol is typically associated with subsequent calorie intake in stress protocols that might induce an obese-like phenotype. It has been demonstrated that cold stress increases CORT secretion in rodents and that CORT secretion is closely associated with the upregulation of hypothalamic orexigenic peptides (such as NPY/AgRP). When cold stress and a high-fat diet are present, NPY and its receptor (Y2R) appear to significantly increase in white adipose tissue. Additionally, it was hypothesized that these modifications would cause adipocytes to proliferate and differentiate, leading to obesity and a state that resembled the metabolic syndrome [156]. Following subordination stress, similar elevations in Y2R have been seen in white adipose tissue [132].

2. Conclusion And Future Perspectives

The major focus of this study is on the impact of stressful environments on the occurrence of obesity and other similar metabolic diseases. According to studies, people eat more when they are stressed as compared to when they are happy because stress causes our bodies to release the hormone cortisol. However, stress-induced eating might be difficult to resist since eating is pleasant. The hormone leptin is important to the regulation of body weight and homeostasis, and leptin resistance is now known as the major contributing factor to obesity. In the brain, specifically in the hypothalamus, leptin promotes weight reduction predominantly through activating the LEPRb pathways. However, it is unknown how the many LEPRb pathway branches function specifically and/or in coordination to control various aspects of food intake behavior and energy expenditure.

In relation to obesity, the interaction between neuronal and hormonal communication through the gutbrain axis has a variety of physiologic aspects, including homeostasis, and might be used as a target for therapy in the future. The precise monitoring of homeostasisand consumption of food might help to produce a more individualized therapy for obesity, regardless of whether it is pharmacological or psychological. This way of treatment quite helpful in the society to control obesity associated diseases.

3. REFERENCE

- [1] WHO. Obesity and overweight: fact sheet World Health Organisation. retrieved on May 19 from http://www.who. int/mediacentre/factsheets/fs311/en/; 2015
- [2] Rasmussen BA, Breen DM, Lam TKT. Lipid sensing in the gut, brain and liver. Trends EndocrinolMetab 2012; 23:49–55
- [3] Flegal KM, Carroll MD, Ogden CL, and Curtin LR. Prevalence and trends in obesity among US adults, 1999- 2008. JAMA 2010;303(3):235-41.
- [4] Peeters A, Barendregt JJ, Willekens F, Mackenbach JP, Al Mamun A, Bonneux L, et al. Obesity in adulthood and its consequences for life expectancy: a life-table analysis. Ann Intern Med 2003;138(1):24-32.
- [5] Aguilera G. Regulation of pituitary ACTH secretion during chronic stress. Front Neuroendocrinol 1994;15(4):321-50.

- [6] Dinneen S, Alzaid A, Miles J, and Rizza R. Metabolic effects of the nocturnal rise in cortisol on carbohydrate metabolism in normal humans. J Clin Invest 1993; 92(5): 2283–2290.
- [7] Rizza RA, Mandarino LJ, and Gerich JE. Cortisol-induced insulin resistance in man: impaired suppression of glucose production and stimulation of glucose utilization due to a postreceptor detect of insulin action. J ClinEndocrinolMetab 1982;54(1):131-8.
- [8] Mangos GJ, Walker BR, Kelly JJ, Lawson JA, Webb DJ, and Whitworth JA. Cortisol inhibits cholinergic vasodilation in the human forearm. Am J Hypertens 2000;13(11):1155-60.
- [9] Huh JY, Park J, Kim JI, Park YJ, Lee YK, Kim JB Kim JB. Deletion of CD1d in adipocytes aggravates adipose tissue inflammation and insulin resistance in obesity. Diabetes 2017;66(4):835-847.
- [10] Selovic A, Sarac J, Missoni S. Changes in adipose tissue distribution during pregnancy estimated by ultrasonography. J MaternFetal Neonatal Med 2016;29(13):2131-7.
- [11] Trouwborst I, Bowser S, Goossens GH, Blaak EE. Ectopic fat accumulation in distinct insulin resistant phenotypes; targets for personalized nutritional interventions. Front Nutr 2018; 5:77
- [12] Martos-Moreno GA, Gil-Campos M, Bueno G, Bahillo P, Bernal S, Feliu A, et al. Obesity associated metabolic impairment is evident at early ages: spanish collaborative study. NutrHosp 2014;30(4):787-93.
- [13] Li X, Zhao Y, Jin Y, Zhang T, Chang X, Liao S, et al. Associations between serum adipocytokines and glycemic tolerance biomarkers in a rural Chinese population. PLoS One 2017;12(8): e0182273.
- [14] Leoni MC, Valsecchi C, Mantelli M, Marastoni L, Tinelli C, Marchi A, et al. Impact of child obesity on adipose tissue physiology: assessment of adipocytokines and inflammatory cytokines as biomarkers of obesity. Pediatr Rep 2010;2(2): e19.
- [15] Dimova R, Tankova T. The role of vaspin in the development of metabolic and glucose tolerance disorders and atherosclerosis. Biomed Res Int 2015; 2015;82348.
- [16] Norseen J, Hosooka T, Hammarstedt A, Yore MM, Kant S, Aryal P, et al. Retinol-binding protein 4 inhibits insulin signaling in adipocytes by inducing proinflammatory cytokines in macrophages through a c-Jun N-terminal kinase- and toll-like receptor 4-dependent and retinol-independent mechanism. Mol Cell Biol 2012;32(10):2010-9.
- [17] Mauro CR, Nguyen BT, Yu P, Tao M, Gao I, Seidman MA, et al. Inflammatory "adiposopathy" in major amputation patients. Ann VascSurg 2013;27(3):346-52.
- [18] Haider A, Yassin A, Doros G, Saad F. Effects of long-term testosterone therapy on patients with "diabesity": results of observational studies of pooled analyses in obese hypogonadal men with type 2 diabetes, Int J Endocrinol 2014;2014:683515.
- [19] Jelenik T, Séquaris G, Kaul K, Ouwens DM, Phielix E, Kotzka J, et al. Tissue-specific differences in the development of insulin resistance in a mouse model for type 1 diabetes. Diabetes 2014;63(11):3856-67.
- [20] Cheung BM, Li C. Diabetes and hypertension: is there a common metabolic pathway? CurrAtheroscler Rep 2012;14(2):160-6.
- [21] Votruba SB, Jensen MD. Regional fat distribution as a factor in FFA metabolism. Annu Rev Nutr2007; 27:149–63.
- [22] Batterham RL, Heffron H, Kapoor S, Chivers JE, Chandarana K, Herzog H, et al. Critical role for peptide YY in protein-mediated satiation and body-weight regulation. Cell Metab 2006;4(3):223–33.

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- [23] Korner J, Inabnet W, Conwell IM, Taveras C, Daud A, Olivero-Rivera L, et al. Differential effects of gastric bypass and banding on circulating gut hormone and leptin levels. Obesity 2006;14(9):1553–61.
- [24] Chandarana K, Drew ME, Emmanuel J, Karra E, Gelegen C, Chan P, et al. Subject Standardization, acclimatisation, and sample processing affect gut hormone levels and appetite in humans. Gastroenterology 2009;136(7):2115–26.
- [25] Batterham RL, Cowley MA, Small CJ, Herzog H, Cohen MA, Dakin CL, et al. Gut hormone PYY3-36 physiologically inhibits food intake. Nature 2002;418(6898):650–4.
- [26] Koegler FH, Enriori PJ, Billes SK, Takahashi DL, Martin MS, Clark RL, et al. Peptide YY(3-36) inhibits morning, but not evening, food intake and decreases body weight in rhesus macaques. Diabetes 2005;54(11):3198–204.
- [27] Neary MT, Batterham RL. Gut hormones: implication for the treatment of obesity. PharmacolTher 2009;124(1):44–56.
- [28] Berglund MM, Hipskind PA, Gehlert DR. Recent developments in our understanding of the physiological role of PPfold peptide receptor subtypes. ExpBiol Med (Maywood) 2003;228(3):217–44.
- [29] Lassmann V, Vague P, Vialettes B, Simon MC. Low plasma levels of pancreatic polypeptide in obesity. Diabetes 1980; 29(6):428–30.
- [30] Glaser B, Zoghlin G, Pienta K, Vinik AI. Pancreatic polypeptide response to secretin in obesity: effects of glucose intolerance. Horm Metab Res 1988;20(5):288–92.
- [31] Batterham RL, Le Roux CW, Cohen MA, Park AJ, Ellis SM, Patterson M, et al. Pancreatic polypeptide reduces appetite and food intake in humans. J ClinEndocrinolMetab 2003; 88(8):3989–92.
- [32] Wisen O, Bjorvell H, Cantor P, Johansson C, Theodorsson E. Plasma concentrations of regulatory peptides in obesity following modified sham feeding (MSF) and a liquid test meal. RegulPept 1992;39(1):43–54.
- [33] Tolhurst G, Reimann F, Gribble FM. Nutritional regulation of glucagon-like peptide-1 secretion. J Physiol2009; 587:27–32.
- [34] Baggio LL, Huang Q, Brown TJ, Drucker DJ. Oxyntomodulin and glucagon-like peptide-1 differentially regulate murine food intake and energy expenditure. Gastroenterology 2004; 127(2):546–58.
- [35] Naslund E, Bogefors J, Skogar S, Gryback P, Jacobsson H, Holst JJ, et al. GLP-1 slows solid gastric emptying and inhibits insulin, glucagon, and PYY release in humans. Am J Physiol 1999;277: R910–6.
- [36] Turton MD, O'Shea D, Gunn I, Beak SA, Edwards CM, Meeran K, et al. A role for glucagon-like peptide-1 in the central regulation of feeding. Nature 1999;379(6560):69–72.
- [37] Verdich C, Flint A, Gutzwiller JP, Naslund E, Beglinger C, Hellstrom PM, et al. A meta-analysis of the effect of glucagon-like peptide-1 (7-36) amide on ad libitum energy intake in humans. J ClinEndocrinolMetab 2001;86(9):4382–9.
- [38] Rehfeld JF, Sun G, Christensen T, Hillingso JG. The predominant cholecystokinin in human plasma and intestine is cholecystokinin-33. J ClinEndocrinolMetab 2001;86(1): 251–8.
- [39] Kopin AS, Mathes WF, McBride EW, Nguyen M, Al-Haider W, Schmitz F, et al. The cholecystokinin-A receptor mediates inhibition of food intake yet is not essential for the maintenance of body weight. J Clin Invest 1999;103(3): 383–91.
- [40] Zwirska-Korczala K, Konturek SJ, Sodowski M, Wylezol M, Kuka D, Sowa P, et al. Basal and postprandial plasma levels of PYY, ghrelin, cholecystokinin, gastrin and insulin in

- women with moderate and morbid obesity and metabolic syndrome. J PhysiolPharmacol2007; 58:13–35.
- [41] Dornonville De La Cour C, Bjorkqvist M, Sandvik AK, Bakke I, Zhao CM, Chen D, et al. Alike cells in the rat stomach contain ghrelin and do not operate under gastrin control. RegulPept 2001;99(2-3):141-50.
- [42] Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K., et al. A role for ghrelin in the central regulation of feeding. Nature 2001;409(6817):194-8.
- [43] Currie PJ, Mirza A, Fuld R, Park D, and Vasselli JR. Ghrelin is an orexigenic and metabolic signaling peptide in the arcuate and paraventricular nuclei. Am J PhysiolRegulIntegr Comp Physiol 2005;289(2): R353-R358
- [44] Cowley MA, Smith RG, Diano S, Tschop M, Pronchuk N, Grove KL, et al. The distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. Neuron 2003;37(4):649-61.
- [45] Seoane LM, Lopez M, Tovar S, Casanueva FF, Senaris R, and Dieguez C. Agoutirelated peptide, neuropeptide Y, and somatostatin-producing neurons are targets for ghrelin actions in the rat hypothalamus. Endocrinology 2003;144(2):544-51
- [46] Abizaid A, Liu ZW, Andrews ZB, Shanabrough M, Borok E, Elsworth JD, et al. Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. J Clin Invest 2006;116(12):3229-39
- [47] Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE, Weigle DS. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. Diabetes 2008; 50(8):1714–9
- [48] Banks WA, Tschop M, Robinson SM, Heiman ML. Extent and direction of ghrelin transport across the blood-brain barrier is determined by its unique primary structure. J PharmacolExpTher 2002;302(2):822–7.
- [49] Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H, Kangawa K, et al. A role for ghrelin in the central regulation of feeding. Nature 2001;409(6817):194–8.
- [50] Williams DL, Grill HJ, Cummings DE, Kaplan JM. Vagotomy dissociates short- and long-term controls of circulating ghrelin. Endocrinology 2003;144(12):5184–7.
- [51] Cabral A, Suescun O, Zigman JM, and Perello M. Ghrelin indirectly activates hypophysiotropic CRF neurons in rodents. PLoS One 2012;7(2): e31462.
- [52] Wren AM, Seal LJ, Cohen MA, Brynes AE, Frost GS, Murphy KG, et al. Ghrelin enhances appetite and increases food intake in humans. J ClinEndocrinolMetab 2001;86(12): 5992–5.
- [53] Feigerlova E, Diene G, Conte-Auriol F, Molinas C, Gennero I, Salles JP, et al. Hyperghrelinemia precedes obesity in Prader–Willi syndrome. J ClinEndocrinolMetab 2008;93(7): 2800–5.
- [54] Friedman JM. The function of leptin in nutrition, weight, and physiology. Nutr Rev 2002;60(10 Pt 2):S1-14; discussion S68-84, 85-7
- [55] Elias CF, Lee C, Kelly J, Aschkenasi C, Ahima RS, Couceyro PR, et al. Leptin activates hypothalamic CART neurons projecting to the spinal cord. Neuron 1998;21(6):1375-85
- [56] Cowley MA, Smart JL, Rubinstein M, Cerdan MG, Diano S, Horvath TL, et al. Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. Nature 2001;411(6836):480-4
- [57] Zakrzewska KE, Cusin I, Stricker-Krongrad A, Boss O, Ricquier D, Jeanrenaud B, et al. Induction of obesity and hyperleptinemia by central glucocorticoid infusion in the rat. Diabetes 1999;48(2):365-70.

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- [58] Zakrzewska KE, Sainsbury A, Cusin I, Rouru J, Jeanrenaud B, and Rohner-Jeanrenaud F. Selective dependence of intracerebroventricular neuropeptide Y-elicited effects on central glucocorticoids. Endocrinology 1999;140(7):3183-7
- [59] Weaver JU, Kopelman PG, McLoughlin L, Forsling ML, and Grossman A. Hyperactivity of the hypothalamopituitary-adrenal axis in obesity: a study of ACTH, AVP, betalipotrophin and cortisol responses to insulin-induced hypoglycaemia. ClinEndocrinol (Oxf.) 1993;39(3):345-50
- [60] Castonguay TW. Glucocorticoids as modulators in the control of feeding. Brain Res Bull 1991;27(3-4):423-8
- [61] Cavagnini F, Croci M, Putignano P, Petroni ML, and Invitti, C. Glucocorticoids and neuroendocrine function. Int J Obes. RelatMetabDisord 2000;24Suppl2: S77-9.
- [62] Savontaus E, Conwell IM, and Wardlaw SL. Effects of adrenalectomy on AGRP, POMC, NPY and CART gene expression in the basal hypothalamus of fed and fasted rats. Brain Res 2002;958(1):130-8
- [63] Morimoto M, Morita N, Ozawa H, Yokoyama K, and Kawata M. Distribution of glucocorticoid receptor immunoreactivity and mRNA in the rat brain: an immunohistochemical and in situ hybridization study. Neurosci Res 1996;26(3):235-69.
- [64] Germano CM, De Castro M, Rorato R, Laguna MT, Antunes-Rodrigues J, Elias CF, et al. Time course effects of adrenalectomy and food intake on cocaine- and amphetamineregulated transcript expression in the hypothalamus. Brain Res 2007; 1166:55-64.
- [65] Green PK, Wilkinson CW, and Woods SC. Intraventricular corticosterone increases the rate of body weight gain in underweight adrenalectomized rats. Endocrinology 1992;130(1):269-75.
- [66] Jacobson L. Glucocorticoid replacement, but not corticotropin-releasing hormone deficiency, prevents adrenalectomy-induced anorexia in mice. Endocrinology 1999;140(1):310-7
- [67] Brindley DN, and Rolland Y. Possible connections between stress, diabetes, obesity, hypertension and altered lipoprotein metabolism that may result in atherosclerosis. ClinSci (Lond) 1989;77(5):453-61.
- [68] La Fleur SE. The effects of glucocorticoids on feeding behavior in rats. PhysiolBehav 2006;89(1):110-4.
- [69] Jahng JW, Kim NY, Ryu V, Yoo SB, Kim BT, Kang DW, et al. Dexamethasone reduces food intake, weight gain and the hypothalamic 5-HT concentration and increases plasma leptin in rats. Eur J Pharmacol 2008;581(1-2):64-70
- [70] Marti O, Marti J, and Armario A. Effects of chronic stress on food intake in rats: influence of stressor intensity and duration of daily exposure. PhysiolBehav 1994;55(4):747-53
- [71] Harris RB, Zhou J, Youngblood BD, Rybkin II, Smagin GN, Ryan DH. Effect of repeated stress on body weight and body composition of rats fed low- and high-fat diets. Am J Physiol 1998;275(6): R1928-38.
- [72] Valles A, Marti O, Garcia A, and Armario A. Single exposure to stressors causes long-lasting, stress-dependent reduction of food intake in rats. Am J PhysiolRegulIntegr Comp Physiol 2000;279(3): R1138-44.
- [73] Baumann H, Morella KK, White DW, Dembski M, Bailon PS, Kim H, et al. The full-length leptin receptor has signaling capabilities of interleukin 6-type cytokine receptors. Proc Natl AcadSci USA 1996; 93(16): 8374–8378
- [74] Chen H, Charlat O, Tartaglia LA, Woolf EA, Weng X, Ellis SJ, et al. Evidence that the diabetes gene encodes the leptin receptor: identification of a mutation in the leptin receptor gene in db/db mice. Cell 1996; 84(3): 491–495

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- [75] Taga T, Kishimoto T. Gp130 and the interleukin-6 family of cytokines. Annu Rev Immunol 1997; 15(1): 797–819
- [76] Morris DL, Rui L. Recent advances in understanding leptin signaling and leptin resistance. Am J PhysiolEndocrinolMetab 2009; 297(6): E1247–E1259
- [77] Banks AS, Davis SM, Bates SH, Myers MG Jr. Activation of downstream signals by the long form of the leptin receptor. J BiolChem 2000; 275(19): 14563–14572
- [78] Tartaglia LA. The leptin receptor. J BiolChem 1997; 272(10): 6093–6096
- [79] Hekerman P, Zeidler J, Bamberg-Lemper S, Knobelspies H, Lavens D, Tavernier J, et al. Pleiotropy of leptin receptor signalling is defined by distinct roles of the intracellular tyrosines. FEBS J 2005; 272(1): 109–119
- [80] White DW, Kuropatwinski KK, Devos R, Baumann H, Tartaglia LA. Leptin receptor (OB-R) signaling. Cytoplasmic domain mutational analysis and evidence for receptor homo-oligomerization. J Biol Chem 1997; 272(7): 4065–4071
- [81] Vaisse C, Halaas JL, Horvath CM, Darnell JE Jr, Stoffel M, Friedman JM. Leptin activation of Stat3 in the hypothalamus of wild-type and ob/ob mice but not db/db mice. Nat Genet 1996; 14 (1): 95–97
- [82] Xu AW, Ste-Marie L, Kaelin CB, Barsh GS. Inactivation of signal transducer and activator of transcription 3 in proopiomelanocortin (Pomc) neurons causes decreased pomc expression, mild obesity, and defects in compensatory refeeding. Endocrinology 2007; 148 (1): 72–80
- [83] Bates SH, Stearns WH, Dundon TA, Schubert M, Tso AW, Wang Y, et al. STAT3 signalling is required for leptin regulation of energy balance but not reproduction. Nature 2003; 421(6925): 856–859
- [84] Jiang L, You J, Yu X, Gonzalez L, Yu Y, Wang Q, et al. Tyrosine-dependent and-independent actions of leptin receptor in control of energy balance and glucose homeostasis. Proc Natl AcadSci USA 2008; 105(47): 18619–18624
- [85] Mütze J, Roth J, Gerstberger R, Hübschle T. Nuclear translocation of the transcription factor STAT5 in the rat brain after systemic leptin administration. Neurosci Lett 2007; 417(3): 286–291
- [86] Gong Y, Ishida-Takahashi R, Villanueva EC, Fingar DC, Münzberg H, Myers MG Jr. The long form of the leptin receptor regulates STAT5 and ribosomal protein S6 via alternate mechanisms. J BiolChem 2007; 282(42): 31019–31027
- [87] Lee JY, Muenzberg H, Gavrilova O, Reed JA, Berryman D, Villanueva EC, et al. Loss of cytokine-STAT5 signaling in the CNS and pituitary gland alters energy balance and leads to obesity. PLoS One 2008; 3(2): e1639
- [88] Bjørbaek C, Buchholz RM, Davis SM, Bates SH, Pierroz DD, Gu H, et al. Divergent roles of SHP-2 in ERK activation by leptin receptors. J Biol Chem 2001; 276(7): 4747–4755
- [89] Rahmouni K, Sigmund CD, Haynes WG, Mark AL. Hypothalamic ERK mediates the anorectic and thermogenic sympathetic effects of leptin. Diabetes 2009; 58(3): 536–542
- [90] Li C, Friedman JM. Leptin receptor activation of SH2 domain containing protein tyrosine phosphatase 2 modulates Ob receptor signal transduction. Proc Natl AcadSci USA 1999; 96(17): 9677–9682
- [91] He Z, Zhang SS, Meng Q, Li S, Zhu HH, Raquil MA, et al. Shp2 controls female body weight and energy balance by integrating leptin and estrogen signals. Mol Cell Biol 2012; 32(10): 1867–1878
- [92] Zhang EE, Chapeau E, Hagihara K, Feng GS. Neuronal Shp2 tyrosine phosphatase controls energy balance and metabolism. Proc Natl AcadSci USA 2004; 101(45): 16064–16069

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- [93] Bjorbak C, Lavery HJ, Bates SH, Olson RK, Davis SM, Flier JS, et al. SOCS3 mediates feedback inhibition of the leptin receptor via Tyr985. J BiolChem 2000; 275(51): 40649– 40657
- [94] You J, Yu Y, Jiang L, Li W, Yu X, Gonzalez L, et al. Signaling through Tyr985 of leptin receptor as an age/diet-dependent switch in the regulation of energy balance. Mol Cell Biol 2010; 30(7): 1650–1659
- [95] Björnholm M, Münzberg H, Leshan RL, Villanueva EC, Bates SH, Louis GW, et al. Mice lacking inhibitory leptin receptor signals are lean with normal endocrine function. J Clin Invest 2007; 117(5): 1354–1360
- [96] Niswender KD, Morton GJ, Stearns WH, Rhodes CJ, Myers MG Jr, Schwartz MW. Intracellular signalling. Key enzyme in leptin-induced anorexia. Nature 2001; 413(6858): 794–795
- [97] Lin X, Taguchi A, Park S, Kushner JA, Li F, Li Y, et al. Dysregulation of insulin receptor substrate 2 in β cells and brain causes obesity and diabetes. J Clin Invest 2004; 114(7): 908–916
- [98] Kubota N, Terauchi Y, Tobe K, Yano W, Suzuki R, Ueki K, et al. Insulin receptor substrate 2 plays a crucial role in βcells and the hypothalamus. J Clin Invest 2004; 114(7): 917–927
- [99] Duan C, Li M, Rui L. SH2-B promotes insulin receptor substrate 1 (IRS1)- and IRS2-mediated activation of the phosphatidylinositol 3-kinase pathway in response to leptin. J BiolChem 2004; 279 (42): 43684–43691
- [100] Ren D, Li M, Duan C, Rui L. Identification of SH2-B as a key regulator of leptin sensitivity, energy balance, and body weight in mice. Cell Metab 2005; 2(2): 95–104
- [101] Ren D, Zhou Y, Morris D, Li M, Li Z, Rui L. Neuronal SH2B1 is essential for controlling energy and glucose homeostasis. J Clin Invest 2007; 117(2): 397–406
- [102] Kim MS, Pak YK, Jang PG, Namkoong C, Choi YS, Won JC, et al. Role of hypothalamic Foxo1 in the regulation of food intake and energy homeostasis. Nat Neurosci 2006; 9(7): 901–906
- [103] Taniguchi CM, Emanuelli B, Kahn CR. Critical nodes in signalling pathways: insights into insulin action. Nat Rev Mol Cell Biol 2006; 7(2): 85–96
- [104] Kitamura T, Feng Y, Kitamura YI, Chua SC Jr, Xu AW, Barsh GS, et al. Forkhead protein FoxO1 mediates Agrp-dependent effects of leptin on food intake. Nat Med 2006; 12(5): 534–540
- [105] Plum L, Lin HV, Dutia R, Tanaka J, Aizawa KS, Matsumoto M, et al. The obesity susceptibility gene Cpe links FoxO1 signaling in hypothalamic pro-opiomelanocortin neurons with regulation of food intake. Nat Med 2009; 15(10): 1195–1201
- [106] Sadagurski M, Leshan RL, Patterson C, Rozzo A, Kuznetsova A, Skorupski J, et al. IRS2 signaling in LepR-b neurons suppresses FoxO1 to control energy balance independently of leptin action. Cell Metab 2012; 15 (5): 703–712
- [107] Cota D, Proulx K, Smith KA, Kozma SC, Thomas G, Woods SC, et al. Hypothalamic mTORsignaling regulates food intake. Science 2006; 312(5775): 927–930
- [108] Maya-Monteiro CM, Bozza PT. Leptin and mTOR: partners in metabolism and inflammation. Cell Cycle 2008; 7(12): 1713–1717
- [109] Hirosumi J, Tuncman G, Chang L, Görgün CZ, Uysal KT, Maeda K, et al. A central role for JNK in obesity and insulin resistance. Nature 2002;420(6913):333–336.
- [110] Sabio G, Davis RJ. TNF and MAP kinase signalling pathways. SeminImmunol 2014;26(3):237–245.

- [111] Evans JL, Goldfine ID, Maddux BA, Grodsky GM. Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. Endocr Rev 2002;23(5):599–622.
- [112] Xu H, Xiong C, He L, Wu B, Peng L, Cheng Y, et al. Trans-resveratrol attenuates high fatty acid-induced P2X7 receptor expression and IL-6 release in PC12 cells: possible role of P38 MAPK pathway. Inflammation 2015;38 (1):327–337.
- [113] Trop-Steinberg S, Azar Y. AP-1 expression and its clinical relevance in immune disorders and cancer. Am J Med Sci 2017;353 (5):474–483.
- [114] Bumrungpert A, Kalpravidh RW, Chitchumroonchokchai C, Chuang CC, West T, Kennedy A, et al. Xanthones from mangosteen prevent lipopolysaccharide-mediated inflammation and insulin resistance in primary cultures of human adipocytes. J Nutr. 2009;139(6):1185–1191.
- [115] Feng J, Lu S, Ou B, Liu Q, Dai J, Jiet C, al. The role of JNK signaling pathway in obesity-driven insulin resistance. Diabetes MetabSyndrobes 2020; 13:1399-1406.
- [116] Li S, Eguchi N, Lau H, Ichii H. The role of the Nrf2 signaling in obesity and insulin resistance. Int J MolSci 2020;21(18):6973.
- [117] Blanchard RJ, Blanchard DC, Takahashi T, and Kelley MJ. Attack and defensive behaviour in the albino rat. AnimBehav 1977;25(3):622-34.
- [118] Blanchard DC, Spencer RL, Weiss SM, Blanchard RJ, McEwen B, and Sakai RR. Visible burrow system as a model of chronic social stress: behavioral and neuroendocrine correlates. Psychoneuroendocrinology 1995;20(2):117-34
- [119] Tamashiro KL, Hegeman MA, and Sakai RR. Chronic social stress in a changing dietary environment. PhysiolBehav 2006;89(4):536-42
- [120] Blanchard DC, Sakai RR, McEwen B, Weiss SM, and Blanchard RJ. Subordination stress: behavioral, brain, and neuroendocrine correlates. Behav Brain Res 1993;58(1-2):113-21.
- [121] McKittrick CR, Blanchard DC, Blanchard RJ, McEwen BS, and Sakai RR. Serotonin receptor binding in a colony model of chronic social stress. Biol. Psychiatry 1995;37(6):383-93.
- [122] McKittrick CR, Magarinos AM, Blanchard DC, Blanchard RJ, McEwen BS, and Sakai RR. Chronic social stress reduces dendritic arbors in CA3 of hippocampus and decreases binding to serotonin transporter sites. Synapse 2000;36(2):85-94.
- [123] Hardy MP, Sottas CM, Ge R, McKittrick CR, Tamashiro KL, McEwen BS, et al. Trends of reproductive hormones in male rats during psychosocial stress: role of glucocorticoid metabolism in behavioral dominance. BiolReprod 2002;67(6):1750-5
- [124] Tamashiro KL, Nguyen MM, Fujikawa T, Xu T, Yun Ma L, Woods SC, and Sakai RR. Metabolic and endocrine consequences of social stress in a visible burrow system. PhysiolBehav 2004;80(5):683-93
- [125] Ginsburg, B., and Allee, W. C. Some effects of conditioning on social dominance and subordination in inbred strains of mice. PhysiolZool 1942;15(4):485-506.
- [126] Miczek KA. A new test for aggression in rats without aversive stimulation: differential effects of d-amphetamine and cocaine. Psychopharmacology (Berl) 1979;60(3):253-9
- [127] Miczek KA, Thompson ML, and Shuster L. Opioid-like analgesia in defeated mice. Science 1982;215(4539):1520-2.
- [128] Bartolomucci A, Palanza P, Gaspani L, Limiroli E, Panerai AE, Ceresini G, et al. Social status in mice: behavioral, endocrine and immune changes are context dependent. Physiol Behav 2001;73(3):401-10.

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- [129] Haller J, Fuchs E, Halasz J, and Makara GB. Defeat is a major stressor in males while social instability is stressful mainly in females: towards the development of a social stress model in female rats. Brain Res Bull 1999;50(1):33-9.
- [130] Bhatnagar S, and Vining C. Facilitation of hypothalamic-pituitary-adrenal responses to novel stress following repeated social stress using the resident/intruder paradigm. HormBehav 2003;43(1):158-65
- [131] Foster MT, Solomon MB, Huhman KL, and Bartness TJ. Social defeat increases food intake, body mass, and adiposity in Syrian hamsters. Am J PhysiolRegulIntegr Comp Physiol 2006;290(5): R1284-93
- [132] Patterson ZR, Khazall R, Mackay H, Anisman H, and Abizaid A. Central ghrelin signaling mediates the metabolic response of C57BL/6 male mice to chronic social defeat stress. Endocrinology 2013;154(3):1080-91
- [133] Devoino L, Alperina E, and Pavina T. Immunological consequences of the reversal of social status in C57BL/6J mice. Brain BehavImmun 2003;17(1):28-34
- [134] Keeney A, Jessop DS, Harbuz MS, Marsden CA, Hogg S, and Blackburn-Munro RE. Differential effects of acute and chronic social defeat stress on hypothalamic-pituitary-adrenal axis function and hippocampal serotonin release in mice. J. Neuroendocrinol 2006;18(5):330-8.
- [135] Krishnan V, Han MH, Graham DL, Berton O, Renthal W, Russo SJ, et al. Molecular adaptations underlying susceptibility and resistance to social defeat in brain reward regions. Cell 2007;131(2):391-404
- [136] Lutter M, Krishnan V, Russo SJ, Jung S, McClung CA, and Nestler EJ. Orexin signaling mediates the antidepressantlike effect of calorie restriction. J Neurosci 2008;28(12):3071-5.
- [137] Lutter M, Sakata I, Osborne-Lawrence S, Rovinsky SA, Anderson JG, Jung S, et al. The orexigenic hormone ghrelin defends against depressive symptoms of chronic stress. Nat Neurosci 2008:11(7):752-3.
- [138] Chuang JC, Perello M, Sakata I, Osborne-Lawrence S, Savitt JM, Lutter M, et al. Ghrelin mediates stress-induced food-reward behavior in mice. J Clin Invest 2011;121(7):2684-92.
- [139] Willner P, Muscat R, and Papp M. Chronic mild stressinduced anhedonia: a realistic animal model of depression. NeurosciBiobehav Rev 1992;16(4):525-34.
- [140] Willner P. Validity, reliability and utility of the chronic mild stress model of depression: a 10-year review and evaluation. Psychopharmacology (Berl) 1997;134(4):319-29.
- [141] Stohr T, Szuran T, Welzl H, Pliska V, Feldon J, and Pryce CR. Lewis/Fischer rat strain differences in endocrine and behavioural responses to environmental challenge. PharmacolBiochemBehav 2000;67(4):809-19.
- [142] Muscat R, and Willner P. Suppression of sucrose drinking by chronic mild unpredictable stress: a methodological analysis. NeurosciBiobehav Rev 1992;16(4):507-17
- [143] Duncko R, Kiss A, Skultetyova I, Rusnak M, and Jezova D. Corticotropin-releasing hormone mRNA levels in response to chronic mild stress rise in male but not in female rats while tyrosine hydroxylase mRNA levels decrease in both sexes. Psychoneuroendocrinology 2001;26(1):77-89.
- [144] Westenbroek C, Ter Horst GJ, Roos MH, Kuipers SD, Trentani A, and Den Boer JA. Gender-specific effects of social housing in rats after chronic mild stress exposure. Prog. NeuropsychopharmacolBiol Psychiatry 2003;27(1):21-30.

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- [145] Patterson ZR, Ducharme R, Anisman H, and Abizaid A. Altered metabolic and neurochemical responses to chronic unpredictable stressors in ghrelin receptor-deficient mice. Eur J Neurosci 2010;32(4):632-9.
- [146] Greeno CG, and Wing RR. Stress-induced eating. Psychol Bull 1994;115(3):444-64.
- [147] Farias-Silva E, Sampaio-Barros MM, Amaral ME, Carneiro EM, Boschero AC, Grassi-Kassisse DM, et al. Subsensitivity to insulin in adipocytes from rats submitted to foot-shock stress. Can J PhysiolPharmacol 2002;80(8):783-9
- [148] Solomon MB, Foster MT, Bartness TJ, and Huhman KL. Social defeat and footshock increase body mass and adiposity in male Syrian hamsters. Am J PhysiolRegulIntegr Comp Physiol 2007;292(1): R283-90.
- [149] Chagra SL, Zavala JK, Hall MV, and Gosselink KL. Acute and repeated restraint differentially activate or exigenic pathways in the rat hypothalamus. RegulPept 2011;167(1):70-8.
- [150] Tamashiro KL, Hegeman MA, Nguyen MM, Melhorn SJ, Ma LY, Woods SC, et al. Dynamic body weight and body composition changes in response to subordination stress. PhysiolBehav 2007;91(4):440-8
- [151] Haque Z, Akbar N, Yasmin F, Haleem MA, and Haleem DJ. Inhibition of immobilization stress-induced anorexia, behavioral deficits, and plasma corticosterone secretion by injected leptin in rats. Stress 2013;16(3):353-62.
- [152] Wang SX, Chen JX, Yue GX, Bai MH, Kou MJ, and Jin ZY. Xiaoyaosan decoction regulates changes in neuropeptide y and leptin receptor in the rat arcuate nucleus after chronic immobilization stress. Evid Based Complement Alternat Med 2012;2012:381278.
- [153] Marquez C, Belda X, and Armario A. Post-stress recovery of pituitary-adrenal hormones and glucose, but not the response during exposure to the stressor, is a marker of stress intensity in highly stressful situations. Brain Res 2002;926(1-2):181-5.
- [154] Dal-Zotto S, Marti O, Delgado R, and Armario A. Potentiation of glucocorticoid release does not modify the long-term effects of a single exposure to immobilization stress. Psychopharmacology (Berl) 2004;177(1-2):230-7
- [155] Geliebter A, Gibson CD, Hernandez DB, Atalayer D, Kwon A, Lee MI, et al. Plasma cortisol levels in response to a cold pressor test did not predict appetite or ad libitum test meal intake in obese women. Appetite 2012;59(3):956-9
- [156] Kuo LE, Kitlinska JB, Tilan JU, Li L, Baker SB, Johnson MD, et al. Neuropeptide Y acts directly in the periphery on fat tissue and mediates stressinduced obesity and metabolic syndrome. Nat Med 2007;13(7):803-11.